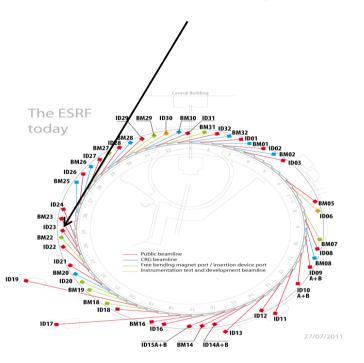
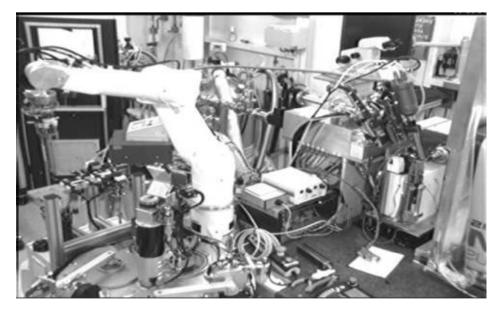
ID23-1 MX beamline news

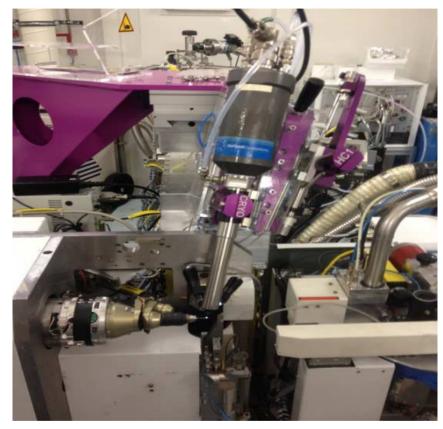


Alexander Popov ESRF, MX group

ID23-1 is a tunable MAD-capable station with a mini-focus X-ray beam which opened to users in 2004 (Nurizzo *et al.*, 2006).



The experimental setup on ID23-1



MD2M mini-diffractometer and Rapid Exchanger (ReX) for automated exchange between HC1 humidifier nozzle and cryo-stream nozzle. Including mechanical support system with alignment axes

FlexHCD sample changer

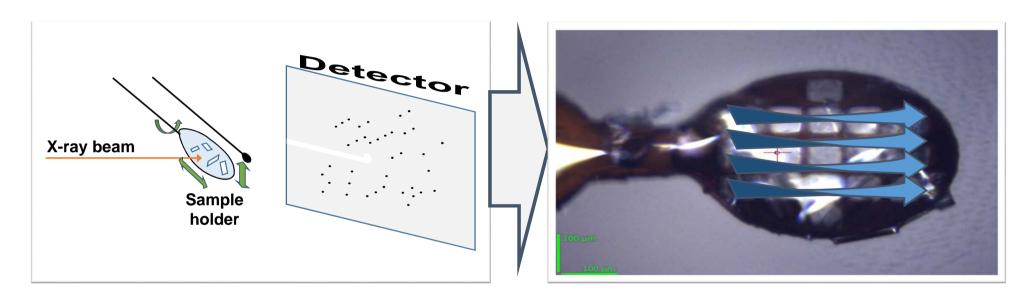


IcePAPS migration

- √ Fast crystal best position screening
- ✓ X-ray centering
- ✓ Crystal cartography
- **✓** Helical Data Collection
- √ X-ray diffraction protein crystal detection

A.Popov 2

X-ray Meshscan – a solid technique for sample analysis

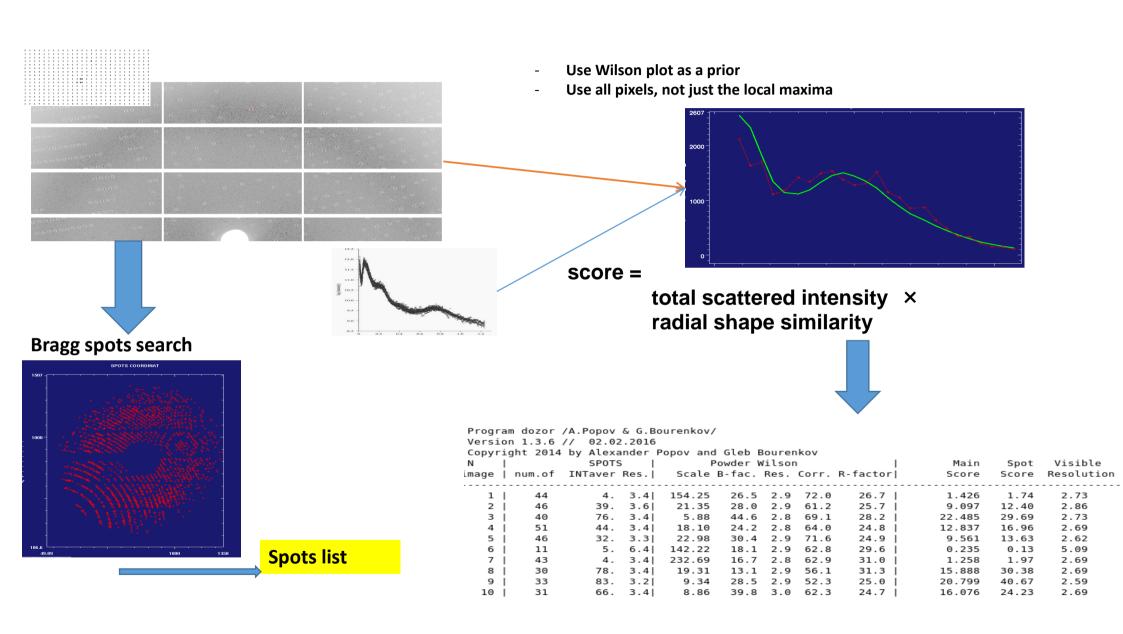


Experimental setup for X-ray crystallography

The data are accumulated during translational movement



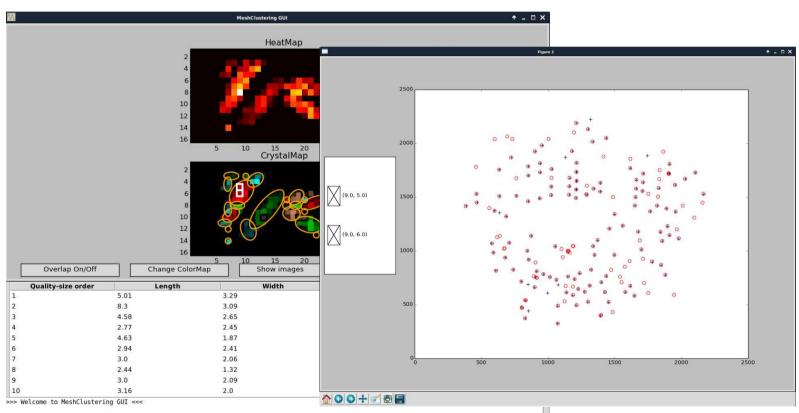
Diffraction images analysis



MeshBest GUI

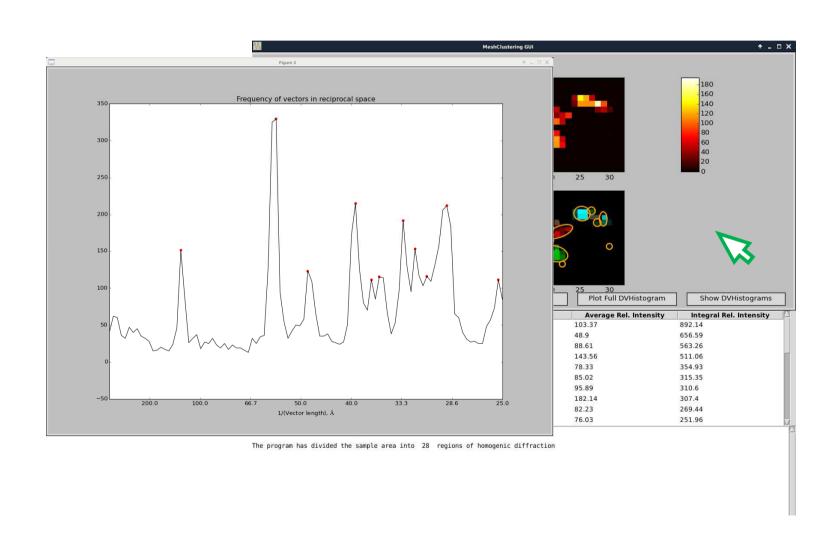
Igor Melnikov

Overview Spot diagram display

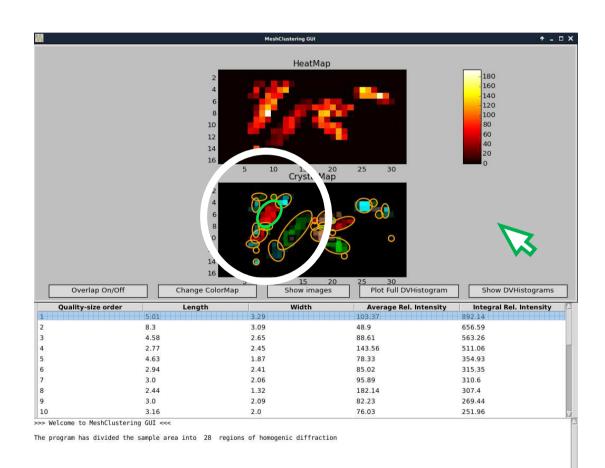


The program has divided the sample area into 28 regions of homogenic diffraction

Interspot distance histogram



Picking the crystal







Gas-sensitive biological crystals processed in pressurized oxygen and krypton atmospheres: deciphering gas channels in proteins using a novel `soak-and-freeze' methodology

B. Lafumat, C. Mueller-Dieckmann, G. Leonard, N. Colloc'h, T. Prangé, T. Giraud, F. Dobias, A. Royant, P. van der Linden and P. Carpentier

A novel cryogenic gas pressure cell has been designed for structural studies of enzymes requiring gaseous substrates. The proof of principle is demonstrated for test crystals. The pressure cell has been designed for the study of O₂-sensitive proteins to reveal pores, channels and reactive centres, and thus to decipher O₂ traffic in proteins.

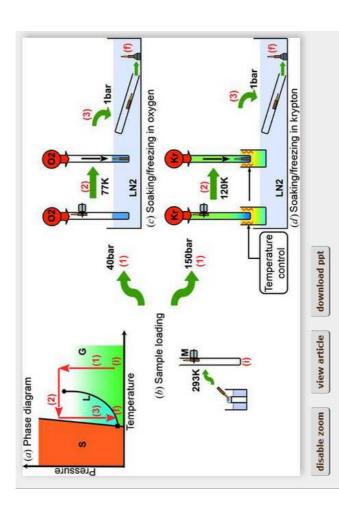
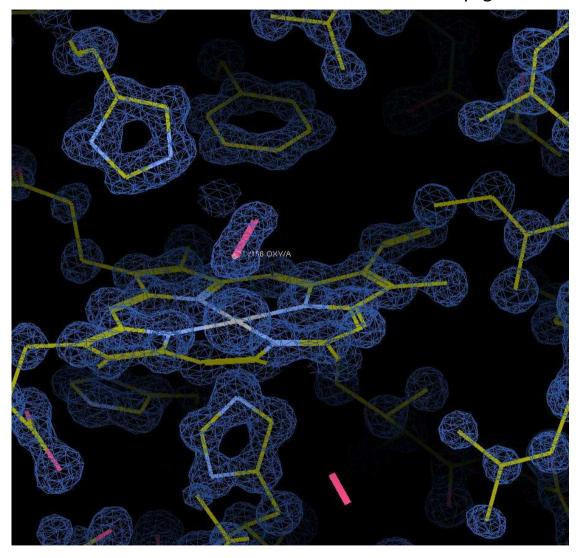
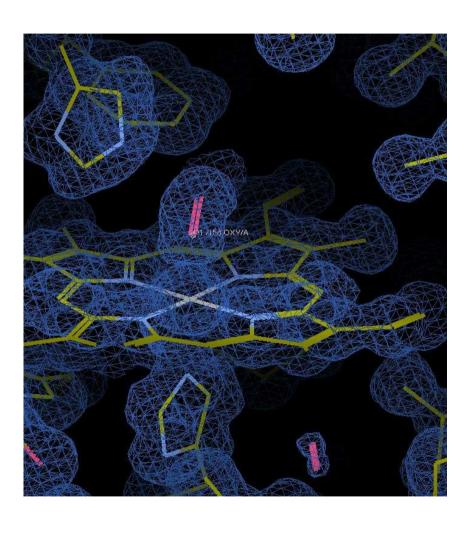


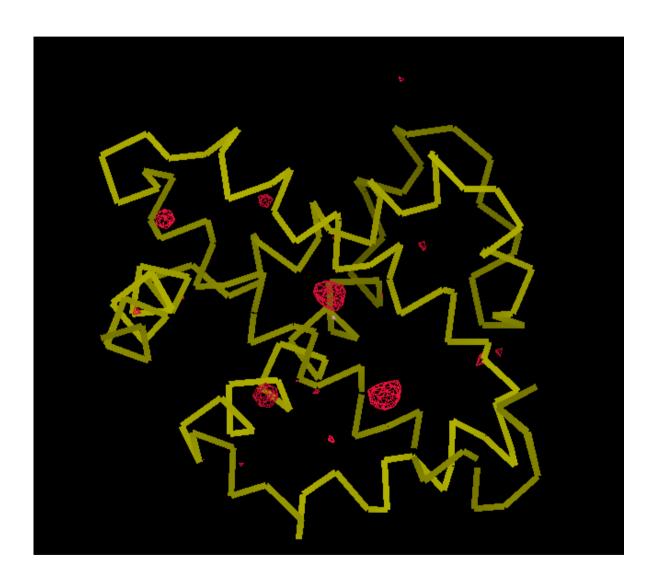
Figure 1
Soak-and-freeze thermodynamics. (a) A typical phase diagram (p, 7), displaying the soak-and-freeze Soak-and-freeze pathway subdivided into its three transformations. (b) Sample fishing from the crystallization tray and loading into the pressure cell. The sample is held at the top of the tube by an external magnet. (c) Soaking and freezing in oxygen. (d) The same process for krypton, with temperature regulation at the bottom of the tube.

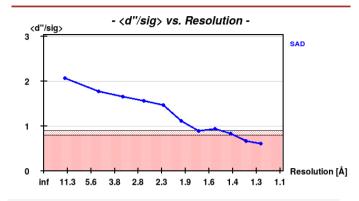
OXY - Myoglobin

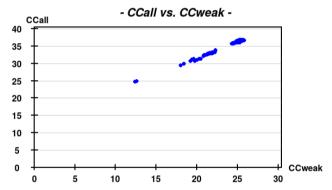


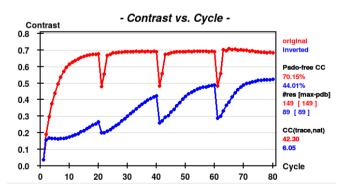


Myoglobin and Kr

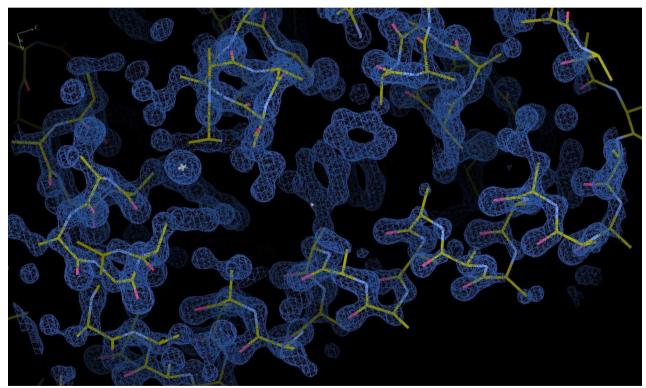








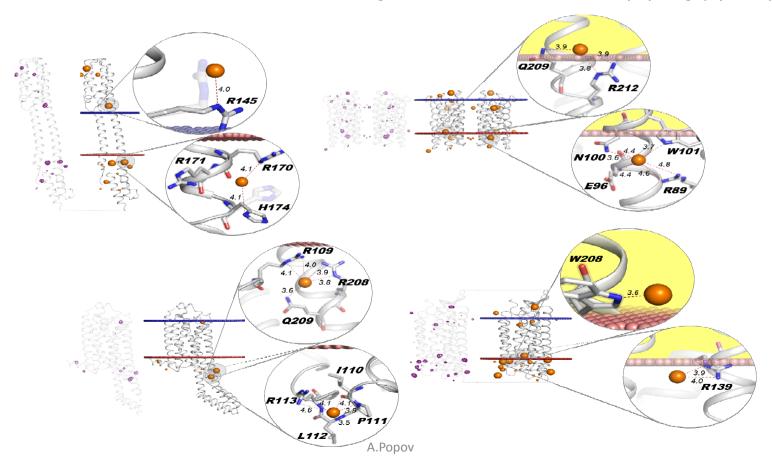
SHELX and Leghemoglobin+Kr



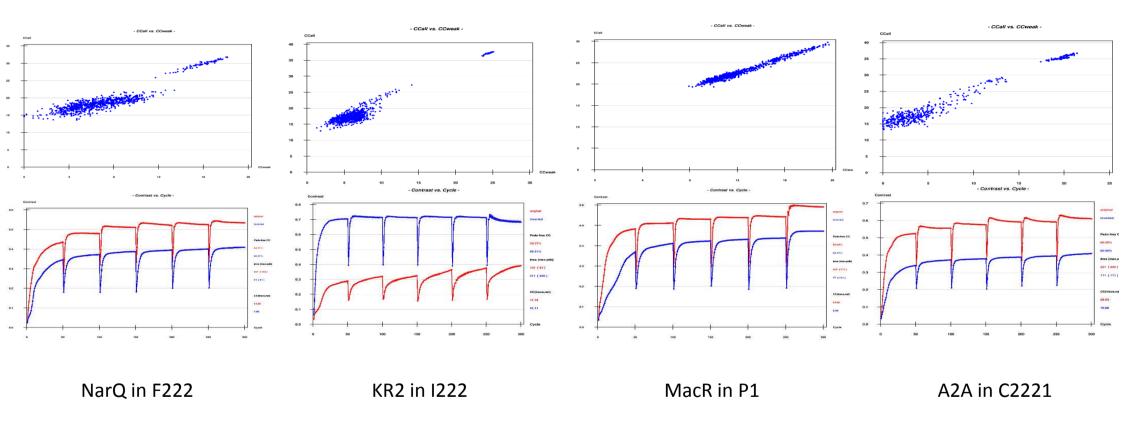
Fast iodide-SAD phasing for high throughput membrane protein structure determination

Igor Melnikov ^a *, Vitaly Polovinkin ^{bce} *, Kirill Kovalev^{ce}, Ivan Gushchin^{ce}, Mikhail Shevtsov^e, Vitaly Shevchenko^{cde}, Alexey Mishin^e, Alexey Alekseev^{ce}, Francisco Rodriguez-Valera^f, Valentin Borshchevskiy^e, Vadim Cherezov^{eg}, Gordon Leonard^a, Valentin Gordeliy^{bce} **, Alexander Popoy^a **

We describe a fast, easy and potentially universal method for solving the crystal structures of membrane proteins via iodide-single-wavelength anomalous diffraction. The potential universality of the method is based on a common feature of membrane proteins – availability at hydrophilic-hydrophobic interface of positively charged residues with which iodide strongly interacts. We show that the method is efficient for different data collection strategies based on either standard or serial X-ray crystallography techniques.

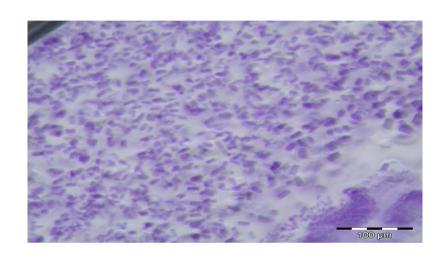


Results (SIRAS)

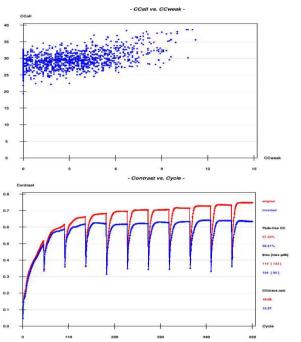


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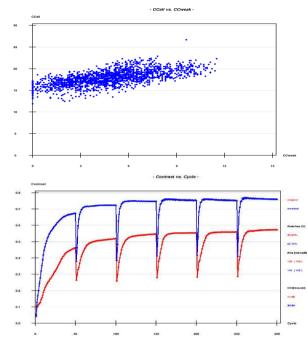
SAD within Mesh&collect pipeline



KR2 small crystals of an average size of $10\text{-}15\mu\text{m}$



SAD, 136 serial datasets merged together



SIRAS, 39 serial datasets merged together + one native dataset from standard collection

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